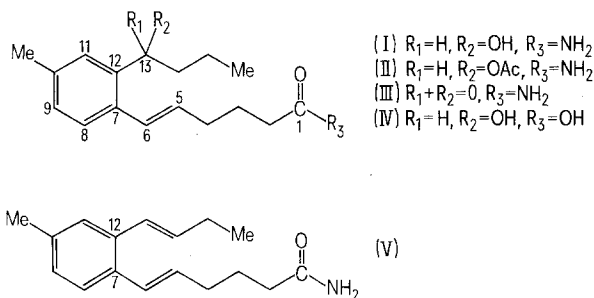


(quintet), respectively, and the relative magnitudes of these shifts indicated that the quintet was due to a CH_2 group β to the amide carbonyl, while the sextet and quartet must be due to CH_2 groups in the other chain.

Finally the aromatic substitution pattern was elucidated. Of the 6 possible positional isomers, only 1 was consistent with the observed order of induced shifts of the aromatic protons in (I), (III) and (V). In compounds (I) and (III), $\text{Eu}(\text{DPM})_3$ associates with both functional groups but the effect of the amide is greater than either of the oxygen functions at C-13. This results in a higher induced shift for the proton at C-8 compared to the other *ortho* proton at C-11 (see Table). In compound (V), only the amide associates with the complex. The induced shift of the proton at C-8 is still the largest but the shifts of the protons at C-9 and C-11 are now identical which is consistent with their equidistance from the amide. Moreover, the low induced shifts of these two protons confirm their *meta* rather than *ortho* substitution relative to the amide chain.



Compounds (I) and (V) probably arise biosynthetically from a suitably unsaturated 10-methylpalmitic acid precursor by a similar pathway (in this case a 7,12 cyclization would be involved) to that discussed recently³ for the antibiotic brefeldin A and the biosynthetically related prostaglandins. We have established⁴ that antibiotic X-537A is assembled from acetate, propionate and butyrate units, suggesting that in the X-537 co-metabolites (I) and (V), the aromatic methyl arises from a propionate unit rather than a C_1 donor system.

Protons at C No.	δ (I)	$\Delta\delta^a$ (I)	δ (III)	$\Delta\delta^a$ (III)	δ (V)	$\Delta\delta^a$ (V)
CH_2	2	2.21 2.35	2.21 2.35	2.25 2.35	2.25 2.35	
CH_2	3	1.77 1.94	1.84 1.91	1.79 1.93		
CH_2	4	2.24 1.10	2.27 1.08	2.25 1.10		
$\text{CH}=\text{}$	5	5.94 0.63	5.89 0.63	5.96 0.58		
$\text{CH}=\text{}$	6	6.68 0.63	6.65 0.62	6.63 0.46		
$\text{CH}=\text{}$	8	7.27 0.28	7.37 0.25	7.25 0.22		
$\text{CH}=\text{}$	9	7.01 0.10	7.20 0.09	6.96 0.08		
$\text{C}-\text{CH}_3$	10	2.31 0.07	2.35 0.06	2.29 0.04		
$\text{CH}=\text{}$	11	7.23 0.23	7.34 0.14	7.25 0.08		
	13	4.95 0.51		6.60 0.21		
	14	1.68 0.33	2.83 0.13	6.06 0.15		
CH_2	15	1.38 0.21	1.68 0.07	2.02 0.06		
CH_3	16	0.92 0.11	0.96 0.02	1.08 0.04		

^a $\Delta\delta$ $\text{Eu}(\text{DPM})_3$ induced paramagnetic shifts.

Zusammenfassung. Die Struktur des *trans*-6-[2-(1-hydroxybutyl)-4-tolyl]-hex-5-enamids (I), eines neuen mikrobiellen Metaboliten, wurde durch Protonenresonanz-Spektroskopie unter Verwendung des Verschiebungsreagenz $\text{Eu}(\text{DPM})_3$ bestimmt.

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³ J. D. BU'LOCK and P. T. CLAY, J. chem. Soc. (D) 1969, 237.

⁴ J. W. WESTLEY, R. H. EVANS JR., D. L. PRUESS and A. STEMPER, J. chem. Soc. (D) 1970, 1467. — J. W. WESTLEY, D. L. PRUESS and R. G. FITCHER, J. chem. Soc. (D) 1972, 161.

The Sterols of the Echinoderm, *Ctenodiscus crispatus* Retzius

Echinoderms have been known for some time to contain mixtures of sterols¹, but recent work^{2,3} has shown these to be more complex than previously envisaged. We report here the composition of the free sterol fraction obtained from the mudstar *Ctenodiscus crispatus* Retzius, Order Phanerozoia, Family Porcellanasteridae. Unlike other sea stars that have been examined, this species is not carnivorous, but feeds by ingesting from muddy ocean bottoms, on which it lives⁴.

Methods. The live animals were blended in chloroform, which was then washed, dried, and chromatographed on a silica-gel column. Visual fractions turned out to contain primarily different classes of metabolites, major ones being neutral glycerides and free sterols. Typically, 87 animals (1425 g net weight) gave 12.1 g of glycerides and 2.72 g of sterols. A clean, crystalline sterol fraction was obtained by chromatography on Florisil (using iso-octane/ether, 3:1) and had m.p. 135–138°, α_D^{25} 0.4 (*c* 1.49 g/100 ml, chloroform). Preparative TLC of the sterol fraction on silica gel (HF₂₅₄+366) containing 20% silver nitrate (4 elutions with chloroform) gave 3 bands, *viz.* Band I (least polar, 85%), Band II (15%) and Band III

(most polar, trace quantity), the first 2 of which corresponded primarily to monoenic and dienic sterols. Band III was not examined further.

Results and discussion. Examination of the GLC and the IR-, UV- and NMR-spectra of the sterol fraction, as well as of its monoacetate and monomethyl ether, suggested the presence of a mixture of cholestenols. The mass spectra (*MS*) of the sterols and of their methyl ethers showed quite clearly that the mixture was primarily a series of homologs of 5 α -cholest-7-en-3-ols^{5,6}. Unusually intense M-2 peaks suggested the presence of dehydrocho-

¹ J. S. GROSSERT, Chem. Soc. Reviews, Lond. 7, 1 (1972).

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³ T. MATSUNO, S. NAGATA and K. MIZUTANI, Nippon Suisan Gakkaishi 38, 144 (1972).

⁴ A. M. D'YAKANOV, Sea Stars (Asteroids) of the USSR Seas (Israel Program for Scientific Translations, Jerusalem, 1968; translated from the Russian, *Morskije zvezdy morei SSSR*), page 16.

⁵ D. R. IDLER, L. M. SAFE and S. SAFE, Steroids 16, 251 (1970).

⁶ B. A. KNIGHTS, J. Gas Chromat. 5, 273 (1967).

GLC-MS data on the sterol mixture^a

Peak Number	Retention time relative to peak a	Abundance, (% of total sterol)	M ⁺ (amu)	Assignment
a	1.00	2.5	388	5 α -cholestan-3 β -ol
			386	uncertain, not cholesterol
			384	uncertain
b	1.12	42.0	386	5 α -cholest-7-en-3 β -ol
c	1.36	8.5	400	24 ξ -methyl-5 α -cholest-7-en-3 β -ol
d	1.43	1.7	—	Uncertain
e	1.67	30.0	414	24 ξ -ethyl-5 α -cholest-7-en-3 β -ol
f	1.95	0.3	428	24 ξ -propyl-5 α -cholest-7-en-3 β -ol
g	1.00	0.3	386	Cholest-5-en-3 β -ol
			384	5 α -cholesta-7,22-dien-3 β -ol
h	1.08	trace	—	Not measured
i	1.20	0.1	398,396	Uncertain, but possibly contains 24 ξ -methyl-5 α -cholesta-7,22-dien-3 β -ol
j	1.32	2.0	398	24-methylene-5 α -cholest-7-en-3 β -ol
k	1.61	0.6	412	Uncertain, but possibly contains 24 ξ -ethyl-5 α -cholesta-7,22-dien-3 β -ol
l	1.73	6.0	412	24-ethylidene-5 α -cholest-7-en-3 β -ol
m	1.87	0.5	426,424	Uncertain
n	2.01	5.5	426	24-propylidene-5 α -cholest-7-en-3 β -ol

^a Glc on 3% OV-101 at 255°C; peaks a-f and g-n correspond to results from bands I and II, respectively.

lestenols, and since the peaks at m/e 388 or 402 were 8% more intense than expected, a cholestanol was also present. Oxidation of the sterol mixture with CrO_3 /acetone, followed by treatment with a trace of base, gave a ketonic material with appropriate spectroscopic properties for 5 α -cholest-7-en-3-ones⁷, although its UV-spectrum indicated the presence of ca. 0.8% cholest-5-en-3 β -ol (cholesterol) in the sterol mixture.

Subjection of the material from Bands I and II to combined GLC-MS confirmed that the separation of Δ^7 monoenes from Δ^7 dienes had been successfully accomplished. Notwithstanding, many GLC peaks still were not homogeneous and consequently complete component identification was not possible. The assignments we have made are shown in the Table. They are based on comparison of the experimentally derived spectra with those reported in the literature⁸ and with those obtained from standard materials⁹. The major peaks b, c, e, j, l and n were easily identified (see Table). Significantly intense $M+2$ ions in the spectra of peaks j, l and n could reasonably be assigned to 24-cholesterol homologs since cholesterol itself has an R_f (in the TLC system used) similar to the R_f of the Δ^7 diene fraction. Peak a contained 5 α -cholestan-3 β -ol from its molecular ion at m/e 388 and its characteristic doublet at m/e 234, 233. The m/e 386 ion in GLC peak a could not have arisen from cholesterol since the m/e 301 ion in the spectrum was not sufficiently intense. The major components in peaks g, i and k possessed parent ions at m/e 384, 398, and 412 respectively and all contained a relatively intense m/e 300 fragment ion. We have tentatively assigned these components respectively to 5 α -cholesta-7,22-dien-3 β -ol and its 24-methyl and 24-ethyl homologs.

The gross composition of this sterol mixture is essentially similar to those studied by GOAD² and SCHEUER¹⁰. The presence of small amounts of 5 α -cholestan-3 β -ol and of Δ^5 -sterols confirms GOAD's findings and current suggestions² for the biosynthesis of sterols in asteroids. Of interest is the presence of several apparently different, previously unreported, C-30 sterols. Only three C-30 cholesterol homologs have been reported to date^{1,11}, two

being Δ^5 and Δ^7 isomers of a sterol with a cyclopropane-containing side chain and the other being (Z)-24-propyldenecholest-5-en-3 β -ol which was found in a local mollusc¹¹. Considering the probable biosynthesis of sterols in sea stars, it is perhaps not unexpected that *C. crispatus* should yield the Δ^7 isomer of this mollusc sterol, together with its 24–28 dihydro derivative. An interesting question raised by these results¹² is whether a comparable mixture of Δ^5 sterols can be isolated from the mud on which these sea stars feed.

Zusammenfassung. Aus dem Schlammstern *Ctenodiscus crispatus* wurde ein kompliziertes Gemisch von Sterolen isoliert, die als Homologe des 5 α -Cholest-7-en-3 β -ols und deren 24-Methylderivate erkannt wurden.

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⁷ The mass spectrum of the ketone mixture differed from that of the parent alcohol by 2 units in all the appropriate peaks.

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⁹ 5 α -cholest-7-en-3 β -ol was obtained from Ikapharm Ltd., Ramat-Gan, Israel, and we thank Professor P. J. SCHEUER for a sample of its 24 ξ -methyl homolog.

¹⁰ K. C. GUPTA and P. J. SCHEUER *Tetrahedron* **24**, 5831 (1967).

¹¹ D. R. IDLER, L. M. SAFE and E. F. MACDONALD, *Steroids* **18**, 545 (1971).

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